



GOVT. ISLAMIA COLLEGE
COOPER ROAD LAHORE

Prebiotic as immunostimulants in aquaculture: A review

Sumama Farooq

Alina Afzal

Mahnoor-ul-Huda

Warda Ishfaq

Soha Hamid

Mehreen Munaf

Hina Mubeen

Department of zoology: Govt. Islamia college for women cooper road

Dr. Mehwish Khan

Significance of study:

we have opted for this topic because the farming and production of finfish and shellfish has become the fastest growing food industry in world and it may help us provide first hand knowledge to the society to improve this field of Pakistan ([Roberfroid et al., 2005](#)) The study provides the practice of intensification that has become common in both finfish and shellfish culture to optimize the returns. Prebiotic termed as immunostimulants are considered as an attractive and promising agent for health and prevention of disease in fish in aquaculture. ([Choque-delgado et al., 2011](#))

Novelty of study :

This review report we have provided several benefits of prebiotics in field of aquaculture. Researchers have already documented the role of prebiotics that they play in the body of host ([Roberfroid et al., 2005](#)). We have gathered information regarding prebiotics I.e they improve water quality , modulation of gut microbiota, increase nutrition, helps in decreasing disease and stress and intensifies immunity and tried to correlate this knowledge with previous findings ([Kocher, 2004](#)).

Gap in knowledge:

The gut microbiota gathered by individuals has proposed to donate to intestinal health as well as disease ([Brown, 2002](#)). Prebiotic supplements focuses on magnifying health by modifying composition of gut microbiota have already become widely available and acceptance is high for it ([Yadav, et al., 2002](#)). However our basic information of gut commensal bacteria variation on a population and its dynamics are still unknown as recent studies has suggested that microbiota adds to obesity, atopic disease, inflammatory bowel diseases and intestinal cancers ([Bron et al., 2012](#)).

Prebiotic as immunostimulants in aquaculture: A review

Abstract

Prebiotics are non nutritious fibers that help grow gut commensal bacteria developing improvements in host's health ([Merrifield et al., 2010](#)). The advantageous effects are because of the byproducts produced during fermentation by gut commensal bacteria ([Ringo et al., 2010](#)). In this report review the undeviating effects of prebiotics on a fish's innate immune system are highlighted. Fructooligosaccharides, Mannan oligosaccharide, inulin and B-glucan are prebiotics that are included in **immunosaccharides** ([Ganguly et al., 2013](#)). These directly intensify innate immune responses by acting at different levels such as: phagocytic, neutrophil activation, activation of alternative complement system, enhanced lysozyme activity. An increase in fish growth and an amelioration of their health status is brought about ([Ringo et al., 2014](#)).

Introduction

Originally prebiotics are non digestible fibers that magnifies the beneficial gut bacteria in its host body ([Smith et al., 1991](#)). It has been seen that the byproducts in fermentation by gut commensal bacteria increase immunity in host ([Abbas et al., 2012](#)). So prebiotics are the basic energy resource for the gut bacteria and called as **functional saccharides**. Another type of functional saccharide further suggested by Kocher called as immunostimulant that prompts the innate immune system directly instead of fermenting by-products of prebiotic. So basically prebiotics are not necessarily an immunostimulant or the other way round. The immunomodulatory activity is stimulated by interactions with their receptors (PRR), such as **B-glucan** and **dectin-1** receptors that are expressed on macrophages ([Beck S et al., 1996](#)). Saccharides might also interact with PRR through microbe associated molecular patterns like teichoic acid, peptidoglycan, glycosylated protein or the polysaccharide of bacteria triggering the innate immune response ([Helliö et al., 2007](#)). Immune response is activated by prebiotics in two ways: **(1)** directly stimulating innate immune system, **(2)** increasing growth of commensal bacteria. This review focuses only on prebiotics that are used in aquaculture and which personally enhance the innate immune system in both finfish and shellfish. The informational details on finfish is gathered in the form of families on behalf of current taxonomical structures, and was discussed by gathering immunostimulant prebiotics into two groups according to chain length: oligosaccharides and polysaccharides ([Alexander JB et al., 1992](#)).

Mechanism of immune response of fish:

In phagocytic leukocytes (monocytes, dendritic cells, macrophages and neutrophils) active host mechanism occur i.e. phagocytosis, in the spleen, head kidney, or other lymphoid organs. There are no statistics on Peyer's patches or patch like lymphoid organs in the intestine of fish. Phagocytosis is the endocytic and phagocytic leukocytes that is the ingesting and engulfing of the cells or particles ([Goethe et al., 1998](#)). It was discovered by measuring the degree of endocytosed zymosan in phagocytic cells the use of microscopy or colorimetric detection. Phagocytosis occurs in a sequence of steps: 1)

microbes are sensed with the help of PRRs such as toll like receptors (TLRs); 2) the microbes engulfed in phagosomes; 3) phagosomes merge with lysosomes, which contain a range of proteases; 4) the microbes are killed via proteolysis. Antigen processing steps followed the antigen presentation to T cells, leads T cells activation and consequently the complete immune device ([Fuller et al., 1998](#)).

Macrophage activation:

Macrophages helps in killing the pathogenic microbes but they also have a closer connections in innate and adaptive immune system to generate maximum immune response. ([Swanson et al., 2000](#)) IFN-g activate the macrophages and direct interactions between MAMPs on bacteria and PRRs on the host cells. Variety of inflammatory cytokines including tumor necrosis factor (TNF), IL-1, IL-12, and others are secreted by activated macrophages. These cytokines are important measures of macrophage activation, they are detected either by PCR or ELISA. ([Teitelbaum et al., 2002](#))

Respiratory burst:

An indication of the oxidative potential of reactive oxygen species is called respiratory burst or oxidative burst, reactive oxygen species i.e. hydrogen peroxide, superoxide anions, and hydroxyl radicals ([Sghir et al., 1998](#)). These species are produced by activated phagocytic cells and they are responsible for killing or destruction of engulfed materials such as microbes. Reactive oxygen species have been extensively used to evaluate the ability of the host to defend towards pathogens. Respiratory bursts from innate immune cells, which includes blood neutrophils, are measured using NBT (nitroblue tetrazolium) or MPO (myeloperoxidase) assays. ([Manning et al., 2004](#)).

Acid phosphatase:

Removal of phosphate groups from phosphorylated molecules by an enzyme is called acid phosphatase. In activated macrophages, acid phosphatase activates the inner pH of macrophage phagolysosomes growing inner acidity. Increase in internal acidity will increase the activation of protease that results in microbial activity. The activity of phosphatase along with macrophage activation, leads to increased phagocytosis and respiratory burst ([Vogt L et al., 2013](#)).

Serum complement activity:

In the immune system, the serum complement system is the non cellular effector response, activated by antigen-specific antibodies, microbial cell surfaces or lectin. Complement is activated through the proteolytic cleavage of precursors to structure new complement units ([Helland B et al., 2008](#)). Some features of complement system are: 1) invading pathogens by forming membrane attack complexes (MAC). (2) Inflammation at local infection site (3) opsonizing invading pathogens by attaching immunoglobulin or complement sub units to their surface that enhances phagocytosis ([Zhang CN et al., 2013](#)).

Phenoloxidase activity:

The status of the innate immune gadget of marine invertebrates such as crayfish, sea cucumber, shrimp or lobster can be measured by Phenoloxidase (PO) activity. The prophenoloxidase (proPO) consists of tyrosinases, catecholases and laccases. PO activity is integral for increasing microbial activity (Soleimani N *et al.*, 2012).

Lysozyme activity:

Lysozyme activity is an enzyme that degrades the peptidoglycan in bacterial walls by hydrolyzing β -(1,4) glycosidic linkages in N-acetylmuramic acid and N-acetylglucosamin. In the mucus, serum, intestine and eggs of marine animals, lysozyme can be determined. Foremost producers of lysozyme are the activated macrophages (Ye JD *et al.*, 2011).

Serum antibody level:

To recognize specific microbial antigens B-lymphocytes produce antibodies. Antibodies neutralize pathogens by binding to their surface antigens and preventing them from attaching to their target cells (Akrami R *et al.*, 2013). Antibodies facilitate phagocytosis of antibody bound pathogens by opsonization (Ai Q *et al.*, 2011).

Hematocrit and leukocyte numbers:

The hematocrit or hemocyte count reflects the total number of cells in the blood i.e. red blood cells, white blood cells and platelets. It can be used as a macroanalysis of the immunological popularity of fish, as the number of immune cells in blood will increase during immune activation. In the blood, the immune cells can be observed as neutrophils, eosinophils, basophils, monocytes and lymphocytes (Dong C *et al.*, 2013).

Oligosaccharides

Fructooligosaccharides (FOS)

FOS are a fast and medium chains of **β -D-fructans** which are evidently present in a lot of factors which includes Jerusalem artichokes, barley, wheat, rye, triticale, banana, garlic, onion, and honey. The **fructosyl units** are certain by way of potential of β -(2-1) glycosidic linkages and connected to a terminal glucose unit. **FOS**, which includes all the non-digestible oligosaccharides composed of fructose and glucose units, are the most frequently studied prebiotics in terrestrial animals and human beings. (Sun Y 2012., *et al.*) Mammalian digestive systems can't hydrolyze β -(2-1) glycosidic linkages due to the fact they lack β -fructosidases. However, remarkable bacteria, which includes lactobacilli and bifidobacteria that do particular β -fructosidases can ferment FOS. Therefore, including FOS in the food graph can selectively enhance the growth and survival of those bacteria within the gastrointestinal (GI) tract of the hosts. (Sohn 2000., *et al.*) **FOS** interacts with TLR2, a membrane bound receptor expressed on macrophages, PMNs (Polymorphonuclear leukocytes or granulocytes), and dendritic cells, which results in immune phone activation through sign transduction pathways (Linehan 2000., *et al.*). A evaluation of the impact of nutritional FOS supplementation as immuno-stimulants is added in Table 1. Readers with similarly pastime on the outcomes of FOS on fish performance, foods usage in fish, fish intestine morphology and microbiota are mentioned the critiques of Ringø and co-authors.

Table 1

Studies the use of fructooligosaccharides as immunostimulants in aquaculture. Abbreviations: ACH50; opportunity supplement activity; ACP, plasma alkaline phosphatase; PO, phenoloxidase; SOD, superoxide dismutase; TCC, overall coelomocyte count; [, boom; /, no change; Y, lower.

Atlantic salmon (200.2±0.6 g)	Dietary 1.0% (10 g kg ⁻¹)	4 months	Whole Blood Neutrophil Oxidative Radical Production /, Serum Lysozyme Activity /
Black amur bream (30.5 ± 0.5 g)	Dietary 0.3 and 0.6%	8 weeks	ACP [, PO [ACH50 [, Ig M [
Caspian roach (0.67±0.03 g)	Dietary 1, 2, 3%	7 weeks	Serum Ig [, Lysozyme activity [, ACH50 [, Resistance to salinity challenge [
Japanese flounder (21 g)	Dietary 0.005% (5.0 g kg ⁻¹)	56 days	Lysozyme activity [, Phagocytic percentage and index /
Red swamp cray fish (15 e 17 g)	Dietary 0.008, 0.01% (8, 10 g kg ⁻¹)	30 days	Immune related genes (crustin1, lysozyme, SOD, and proPO) [, Phagocytic activity [, SOD [, Survival against A. hydrophila [
Sea cucumber (5.06±0.10 g)	Dietary 0.25, 0.5%	8 weeks	0.5% FOS TCC [, Phagocytosis [, PO [, Resistance

			to V. splendidus [
Sea cucumber (3.72±0.16 g)	Dietary 0.4, 0.8, 1.6%	50 days	No significant immunological improvement observed
Stellate sturgeon (30.16±0.14 g)	Dietary 1, 2%	11 weeks	1%FOS; serum lysozyme activity [
Yellow croaker (7.82±0.68 g)	Dietary 0.2, 0.4%	10 weeks	No statistically significant innate immunity improvement observed

Salmonidae

Atlantic salmon (*Salmo salar* L.). Grisdale-Helland and co- authors accomplished a 4-month find out out about feeding Atlantic salmon 10 g FOS kg⁻¹ and published no massive distinction in feed performance or energy retention withinside the fish that were fed FOS. Moreover, there were no proper sized outcomes on blood neutrophil oxidative radical production (NBT) or serum lysozyme endeavor in the FOS nutritional groups. ([Benites 2008., et al](#))

Cyprinidae

Black amur bream (*Megalobrama terminalis*). Three particular doses of FOS (0, 3, or 6 g kg⁻¹) fed for 8 weeks to black amur bream (triangular bream) especially prolonged the plasma alkaline phosphatase (ACP), PO, preference supplement (ACH50) activities, and immunoglobulin M content material. ([Klebaniuk 2008., et al](#)) However, no statistically significant versions were placed in leucocyte counts, entire serum protein, and globulin content. In addition, nutritional FOS did not make more the survival rate in reaction to an *Aeromonas hydrophila* challenge. Caspian roach (*Rutilus rutilus*). In a modern study, the consequences of nutritional FOS including 10, 20, or 30 g kg⁻¹ on caspian roach fry became investigated Immunoglobulin levels, lysozyme activity, and ACH50 have been significantly increased in the organizations fed 20 and 30 g FOS kg⁻¹. ([Yang 2009., et al](#)) Only lysozyme undertaking was extended in the group fed 10 g FOS kg⁻¹. Dietary FOS dietary supplements increased the resistance of the fry to salinity pressure demanding situations irrespective of the supplement level, however entirely the team fed 3% FOS had a drastically better survival rate.. ([Torricellas 2014., et al](#))

Pleuronectiformes

Japanese flounder (*Paralichthys olivaceus*). Japanese flounder fed 5.0 g FOS kg⁻¹ for 56 days were tested for lysozyme activity, the proportion of phagocytic cells that took up a marker (phagocytic percent), and phagocytic index. ([Staykav 2007., et al](#)) FOS administration considerably better lysozyme activity, but no longer the phagocytic percentage or phagocytic index, in comparison to the manage weight-reduction plan group. . ([Rodrigues-Estrade 2009., et al](#)) However, while fed a combination of FOS and MOS (mannanoligosaccharide)(5.0 g kg⁻¹), the flounder proven a marginal boom in phagocytic activity. ([Rodrigues-Estrade 2013., et al](#))

Acipenseridae

8 Stellate sturgeon (*Acipenser stellatus*). Akrami and co-authors performed a studies about to determine the have an impact on of nutritional FOS; 10 and 20 g kg⁻¹, on innate immune responses of stellate sturgeon juveniles in an 11 week trial. ([Samrongpan 2009., et al](#)) Juvenile fish fed 10 g kg⁻¹ FOS-supplemented weight loss plan found out considerably elevated serum lysozyme activity. However, no development became as soon as positioned in the group fed 20 g FOS kg⁻¹. Furthermore, 10 g FOS kg⁻¹ did not considerably have an effect on the respiratory burst undertaking of leucocytes, but the lowest activity was once observed in the 20 g FOS kg⁻¹ fed group, which was even lower than that of the manipulate group.. ([Torrecillas 2011., et al](#))

Salmonidae

Grisdale Hell and et.al analyze the result of dietary MOS supplements by nourishing Atlantic Salmon. The outcome of this work showed no expand in noticed in MOD. Extra studies are required to wind up anyhow, dietary MOS has any result on invate freedom and ailment receptivity. ([Groff 2000., et al](#))

Rainbow trout (*Oncorhynchus mykiss* Walbaum)

External cell wall of s.cerevisial originated thickening production. In earliest case supplement of 2g MOS kg crucially enhanced antigen production. In next case the outcome was less powerful which might revealed a small stability of MOS effects on immune response. Rodrigues Estrada considered that the effects of supplements of MOS in diet of rainbow trout Juvenile fish in a 12 weeks feeding trial. Phagocytic activity and hemolytic act contrasted to fingerlings fed control diet. Two level of MOS learn the result of MOS on freedom. The result revealed that Phagocytes and hematocrit act of fish sustain 5g MOS kg⁻¹ were enhanced as constrast to control fed fish yet no improvement was noticed .It is also noticed that the fish fed had a higher frequency.

Cichlidae

Tilapia(*Oreochomis niloticus*)

B/w control and MOS fed groups the supplements increased acceptance in case of streptococcus aglactia bacteria.

Moronidae

European sea bass(*Dicentrarchus labrax*).

In mucus fish the immune activates and outcome in vivo revelation to *V.anguillarum* was looked into European sea bass sustained that it involved two level of MOS for 8weeks. In stomach mucus dietary MOS enhanced lysozyme act still there was no detectable result on antibody activity. Grouping in vivo revelation, the virus was improved from liver. In fish nourishment MOS pathogen was not found. In proximal intestine and distal intestine the feeding MOS enlarge the number of goblet cells. MOS feeding enlarged the lamina propia, included infiltrated eosinophilic granulocytes. This work indicates useful effects of nutritive MOS on some mechanism which provide instinctual defence. In fish G-1 area is a major disease.

Pleuronectiformes

Japanese flounder. Dietary MOS activated lysozyme act, still no remarkable distinction were noticed for phagocytes proportion and phagocytes indicator as contrast with fish nourishment.

Parastacidal

Marron (*Cherax tenrimanus*). Two research have examined the dietary result of MOS on immune reaction. Sang revealed three stress cases. Marron were nourished with three different levels of MOS to challenge. Before showing the investigation group it was showed that NH₃ was only nourished 4g MOS kg-1 for 42days. In bacterial study the MOS exhibited excessive survival rates. In all fishes the hemocyte were decreased. THCS were excessive in marron than the comparison group. The comparison group also revealed that the THCs were reduced whereas contaminated with microbe or exposed to air. MOS increased granular cells in live transport study and decreased hemolymph-clotting time. MOS enhanced the health of fish. The effect of six different MOS levels was examined on bacillemia and hemolymph. Bacillemia was lower in MOS supply group than the control fed animals. Marron had shorter hemolymph. In supplementary groups Mos is high as contrast to control group.

Crayfish; On immunological parameters the proportion of hyaline cells, the proportion of semi granular cells and bacteremia were looked into crayfish.

Palinuridae

Tropical spiny lobster (*Panulirus ornatus*). Sang examined the outcome of MOS on Juvenile tropical spiny lobsters. At the end of culture period the lobsters were challenged for seven days with vibrio sp. The MOS supply enhanced the survival rate. The bacteria in hemolymph decreases during infection.

Stichopodidae

Sea cucumber. In sea cucumber the dietary MOS increase TCC, phagocytes of colomocytes, super oxide anion production and SOD activity than the control diet. The union of B glucan more strongly enlarged high level of immune parameters as contrast to supplementing diet . It enhanced the

survival rate of sea cucumbers. In sea cucumbers, TCC and superoxide anion formation were high than control group. However no distinction noticed. Sea cucumber had higher survival rate.

Immunogen(An antigen capable of including an immune response).

Cyprinidae

Common carp (Cyprinus carpio L)

Carp were given a diet augmented with immunogen. Immunogen is a trade product carrying two prebiotics, MOS & B-glucan. All amount increase leucocyte counts. The carp nourished Immunogen Weight. The mortality rate was lower. In this class the mortality rate is highest.

Galactooligosaccharide

Lactose consist of 2-20 molecules of galactose and glucose. The prebiotic result has been examined in endothermic animals. Only two studies have analysed.

salmonidae

Atlantic salmon. Atlantic salmon were nourished in a 4month study. In serum Lysozyme act no distinction were noticed. It is a main species in fish farming. The effect on immune parameters should be conducted.

Sciaenidae

Red drum (Sciaenops acellatus)

Juvenile red drums were nourished with GOS for 8weeks still no change in NBT levels. The peak of microvilli in pyloric caeca, proximal and mid intestine were increased as contrast to base diet group.

Arabinoxylan oligosaccharide

The fermentation product which is found in cell wall of many grains is Arabinoxylan. Arabinoxylan are saccharide chains in which X-L arabinofuranose units are attached. Their structural diversity may vary.

Acipenseridae

Siberian sturgeon

Two studies have analyzed the dietary effect of two different AXOS types.

>AXOS - 3 - 0.25

> AXOS - 32 - 0.30

Geraylon showed that both AXOS preparations increased phagocytes activities. The fish serum and respiratory burst activity were not effects affected by feeding.

Polysaccharides

With the great variety in molecular structure monosaccharides are bounded by glycosidic bonds to form long chain of Polysaccharides. Inulin, B-glucan,

and chitin/chitosan are Dietary polysaccharides which have been studied in detail as immunostimulants or prebiotic and many scientist have given review on those extensively studied polysaccharides . Nowadays insulin and B-glucan are widespread consensus among scientist that are classified as prebiotic ([Geralylou et al., 2013](#)). Chitosan and alginate have immunostimulatory activities , according to some studies ,in the host gut their capability to elicit advantageous microbes still remain to be explored . Prebiotic polysaccharides involved in stimulating the fish innate immune system through mechanism are also not identified. However, it is very acceptable that microorganisms developed some prebiotic polysaccharides from cell wall components and MAMPs interacting with specific PRRs expressed on innate immune cells are the functions of them. Innate immune cells become active. ([Zhou et al., 2012](#))

Inulin

Different plants like chicory, cornflower, and dandelion have inulin which is a kind of fructan polysaccharide . When absorbed , the bacteria present in the colon just can digest the polysaccharide. Hydrolysis of insulin can produce FOS as an central artifact . It has been clearly accepted that the optimum dietary dose of inulin in the fish feed 1 g kg¹ can change the immune system of fish. It is not obvious, however, can inulin alone have a function as immunostimulant([Lam et al., 2013](#)). Many previous studies have been focal point that is produced from bacterial inulin fermentation such as butyric acid, propanoic acid, and FOS. Bifidobacteria, lactic acid bacteria (LAB), and clostridia are all known to ferment inulin are called as byproduct .The identification for the specific receptors of insulin should be remained . Inulin have a specific unit which is FOS ,related with TLR2([Chiu et al., 2008](#))

Cichlidae

Nile tilapia. by using Nile tilapia as a fish model, increase occur in hematocrit, NBT activity level, and lysozyme activity by the supply of 5kg of a dietary insulin([Geng et al., 2011](#)) Innate immunity have a stimulatory effect of insulin . In the group there is a statistically important stimulation in NBT activity by feeding insulin for two months. Moreover the adaptation rate following A hydrophila challenge was increased in the inulin fed group .

Acipenseridae

Beluga (Huso huso). For 8 weeks at three different levels 1,2 and 3g in the diet of beluga juveniles inulin was included. Compared to the other inulin dose groups and the control group ,the group fed 1g inulin kg¹ had tremendously increased white blood cellcounts symbolizing stimulation of the immune system ([Harikrishnan et al.,2011](#)).In addition to this innate immune responses such as neutrophil electroactivity NBT tests and phagocytic activity were not experimented in this study.

Sparidae

Gilthead seabream. According to the three studies that we have conducted show the effect of inulin on the immune system of gilt head sea-bream .To study the innate immune response both in vitro and in vivo ,the first study was conducted .In the in vitro experiment, inulin incubated HK leukocytes which were not importantly separate in terms of leucocyte per-oxidase

content and phagocytosis. The in vivo study, the mixing of inulin to the diet also expressed nothing special improvement in per-oxidase activity complement activation, phagocytic ability, or natural cytotoxic activities. In the second study ,by adding 10 g kg¹ inulin was effectively increased serum complement activity , IgM levels, leucocyte phagocytic activity, and leucocyte respiratory burst activity. Moreover ,the gene function was obviously up_ regulated for T cell receptor (TCR) beta, MHC1 alpha, and beta defensin. However, the signers expressing augmentation of the immune system did not correlate with the results of a challenge performance using Photobacterium damsela sub sp piscicida. There were no significant stimulation in the cumulative survival compared to the control.

It was failed to increase innate immune makers by dietary administration of inulin (10 g kg¹) to gilt-head sea bream for 28 days such as natural haemolytic complement activity; leucocyte per-oxidase and serum ; phagocytosis; and cytotoxic activity respiratory burst.

Pimelodidae

Hybrid surubim (Pseudoplatystoma sp.).According to an experiment with hybrid surubim ,when the fish were fed 5 g inulin kg¹ supplement ,there was no significant increase in innate immune parameters .However in the surubim intestine large numbers of 2AB were propagalid ,which might be according to classical description of prebiotic given by Roberfroid .

Table:4

using insulin as imunostimulants in aqua culture, abbreviations, ACH50 : alternative compliment activity;ACP,plasma alkaline phosphate, Gc granular cells ; po,phenoloxidase; SOD, superoxide dimutase , TCC total coelomycetes count; THC total hematocytes count,increase, no change , decrease ([kudrenko et al., 2009](#)).

fish model (weight) Route of administration and dose duration of administration results

beluga great sturgeons 16.14+- 0.38g	dietary 1.0,2.0,3.0% insulin	8 weeks	RBC count total increase□ MCH□	WBC
glith&sea bream 50g	dietary 1.0 (10kg - 1)insulin	2 and 4 weeks	dose dependent alkaline phosphate□ serum compliment activity□, level□ leucocytes phagocytic level□	IgM
hybrid surubim	dietary 0.5% insulin	15 days	lactic acidbacteria□	

(73.6+-19.5 g)			vibrio spp pseudomonas spp total ig
Nile tilapia (11.00+-0.2g)	dietary 0.5%(5gkg-1)insulin	1 and 2 months	hematocrit NBT lysosyme activity high

conclusion :

Fish farming has become one of the greater aquatic industries. In contrast to natural conditions in which fish freely use different types of food, the limitations of food sources available for aquacultured fishes may be limited. One indication of this may be lessened diversity of the intestinal microbial community in farmed fish ([Dhanasiri et al., 2011](#)). Feeding with many dietary additives such as probiotics, prebiotics, vitamins, or crude plant waste may compensate for this process and result in beneficial effects for aquaculture ([Galli et al., 2009](#)). Prebiotics, or immunosaccharides, can also activate the innate immune system directly or by relation with MAMPs that interact with PRRs expressed on immune cells. Proper immune responses are useful not only for combating pathogens, but also for proper weight gain ([Cerezuela et al., 2011](#)). Many studies show that feeding with both prebiotics and immunosaccharides is useful in fish aquacultures of various types of vertebrates and invertebrates ([Ganguly et al., 2010](#)). To better the efficiency of the prebiotics and probiotics used in aquaculture, and monitor more immunostimulatory prebiotics, additional work is provided to enhance the ligand receptor interactions, signal transduction pathways involved in this process and the types of cytokines secreted ([Shoelson et al., 2007](#)). Investigation into the effects of prebiotics and probiotics on the immune system of finfish and crustaceans could be more better or high in their merit if innate immune responses were biologically linked to overall gut health ([Heredia et al., 2012](#)).

Acknowledgments:

The research was the part of the project titled 'Development of A Lactic Acid Bacteria-Based Functional Feed Additive for Antibiotic Free Aquaculture of Flounders' funded by the Ministry of Oceans and Fisheries, ([Gomez et al., 2008](#)). Korea and supported by the National Research Foundation of Korea (NRF) ([Perez et al., 2010](#)).

References:

- [1] [Roberfroid MB. Introducing inulin-type fructans. Brit J Nutr 2005;93\(1\): 13e26.](#)
- [2] [Choque-Delgado GT, Tamashiro WMSC, Marostica Jr MR, Moreno YMF, Pastore GM. The putative effects of prebiotics as](#)

- [immunomodulatory agents. Food Res Int 2011;44:3167e73.](#)
- [3] [Roberfroid M. Dietary fibre, inulin and oligofructose: a review comparing their physiological effects. CRC Crit Rev Food Sci Technol 1993;33:103e48.](#)
- [4] [Kocher A. The potential for immunosaccharides to maximise growth performance: a review of six published meta-analyses on Bio-Mos. In: Tucker LA, Taylor-Pickard JA, editors. Interfacing immunity, gut health and performance. Nottingham University Press; 2004. pp. 107e16.](#)
- [5] [Brown GD, Taylor PR, Reid DM, Willment JA, Williams DL, Martinez Pomares L, et al. Dectin-1 is a major b-glucan receptor on macrophages. J Exp Med 2002;196:407e12.](#)
- [6] [Yadav M, Schorey JS. The b-glucan receptor dectin-1 functions together with TLR2 to mediate macrophage activation by mycobacteria. Blood 2002;108: 3168e75.](#)
- [7] [Bron PA, van Baarlen P, Kleerebezem M. Emerging molecular insights into the interaction between probiotics and the host intestinal mucosa. Nat Rev Microbiol 2012;10:66e78.](#)
- [8] [Merrifield DL, Dimitroglou A, Foey A, Davies SJ, Baker RR, Børgwald J, et al. The current status and future focus of probiotic and prebiotic applications for salmonids. Aquaculture 2010;302:1e18.](#)
- [9] [Ringø E, Olsen RE, Gifstad TØ, Dalmo RA, Amlund H, Hemre G-I, et al. Prebiotics in aquaculture: a review. Aquacult Nutr 2010;16:117e36.](#)
- [10] [Ganguly S, Dora KC, Sarkar S, Chowdhury S. Supplementation of prebiotics in fish feed: a review. Rev Fish Biol Fish 2013;23:195e9.](#)
- [11] [Ringø E, Dimitroglou A, Hoseinifar SH, Davies SJ. Prebiotics in finfish: an up date. In: Merrifield DL, Ringø E, editors. Aquaculture nutrition: gut health, probiotics and prebiotics. Oxford, UK: Wiley-Blackwell Publishing; 2014. pp. 360e400.](#)
- [12] [FishBase version \(12/2013\); 2013](#)
- [13] [Abbas AK, Lichtman AH, Pillai S. Cellular and molecular immunology. 7th ed. PA, USA: Saunders Elsevier Philadelphia; 2012.](#)
- [14] [Smith VJ, Soderhall K. A comparison of phenoloxidase activity in the blood of marine invertebrates. Dev Comp Immunol 1991;15:251e61.](#)
- [15] [Beck G, Habicht GS. Immunity and the invertebrates. Sci Am November 1996: 60e6.](#)
- [16] [Hellio C, Bado-Nilles A, Gagnaire B, Renault T, Thomas-Guyon H. Demonstration of a true phenoloxidase activity and activation of a ProPO cascade in pacific oyster, Crassostrea gigas \(Thunberg\) in vitro. Fish Shellfish Immunol 2007;22:433e40.](#)
- [17] [Alexander JB, Ingram GA. Noncellular nonspecific defense mechanisms of fish. Ann Rev Fish Dis 1992;2:249e80.](#)
- [18] [Goethe R, Phi-van L. Posttranscriptional lipopolysaccharide regulation of the lysozyme gene at processing of the primary transcript in myelomonocytic HD11 cells. J Immunol 1998;160:4970e8.](#)
- [19] [Fuller R, Gibson GR. Probiotics and prebiotics: microflora management for improved gut health. Clin Microbiol Infec 1998;4\(9\):477e80.](#)
- [20] [Swanson KS, Grieshop CM, Flickinger EA, Bauer LL, Wolf BW, Chow J, et al. Fructooligosaccharides and Lactobacillus acidophilus modify bowel function and protein catabolites excreted by healthy humans. J Nutr 132.10:](#)

[3042e3050.](#)

[21] [Teitelbaum JE, Walker WA. Nutritional impact of pre-and probiotics as protective gastrointestinal organisms. Ann Rev Nutr 2002;22\(1\):107e38.](#)

[22] [Sghir A, Chow JM, Mackie RI. Continuous culture selection of bifidobacteria and lactobacilli from human fecal samples using fructooligosaccharides as selective substrate. J Appl Microbiol 1998;85:769e77.](#)

[23] [Manning TS, Gibson GR. Prebiotics. Best Pract Res Clin Ga 2004;18\(2\):287e98.](#)

[24] [Vogt L, Ramasamy U, Meyer D, Pullens G, Venema K, Faas MM, et al. Immune modulation by different types of b2 / 1-fructans is toll-like receptor dependent. PLoS One 2013;8\(7\):e68367.](#)

[25] [Grisdale-Helland B, Helland SJ, Gatlin III DM. The effects of dietary supplementation with mannanoligosaccharide, fructooligosaccharide or galactooligosaccharide on the growth and feed utilization of Atlantic salmon \(*Salmo salar* L.\). Aquaculture 2008;283:163e7.](#)

[26] [Zhang CN, Li XF, Xu WN, Jiang GZ, Lu KL, Wang LN, et al. Combined effects of dietary fructooligosaccharide and *Bacillus licheniformis* on innate immunity, antioxidant capability and disease resistance of triangular bream \(*Megalobrama terminalis*\). Fish Shellfish Immunol 2013;35:1380e6.](#)

[27] [Soleimani N, Hoseinifar SH, Merrifield DL, Barati M, Abadi ZH. Dietary supplementation of fructooligosaccharide \(FOS\) improves the innate immune response, stress resistance, digestive enzyme activities and growth performance of Caspian roach \(*Rutilus rutilus*\) fry. Fish Shellfish Immunol 2012;32\(2\):316e21.](#)

[28] [Ye JD, Wang K, Li FD, Sun YZ. Single or combined effects of fructo- and mannan oligosaccharide supplements and *Bacillus clausii* on the growth, feed utilization, body composition, digestive enzyme activity, innate immune response and lipid metabolism of Japanese flounder *Paralichthys olivaceus*. Aquacult Nutr 2011;17:e902e11.](#)

[29] [Akrami R, Iri Y, Rostami HK, Razeghi Mansour M. Effect of dietary supplementation of fructooligosaccharide \(FOS\) on growth performance, survival, *Lactobacillus* bacterial population and hemato-immunological parameters of stellate sturgeon \(*Acipenser stellatus*\) juvenile. Fish Shellfish Immunol 2013;35:1235e9.](#)

[30] [Ai Q, Xu H, Mai K, Xu W, Wang J, Zhang W. Effects of dietary supplementation of *Bacillus subtilis* and fructooligosaccharide on growth performance, survival, non-specific immune response and disease resistance of juvenile large yellow croaker, *Larimichthys crocea*. Aquaculture 2011;317:155e61.](#)

[31] [Dong C, Wang J. Immunostimulatory effects of dietary fructooligosaccharides on red swamp crayfish, *Procambarus clarkii* \(Girard\). Aquacult Res 2013;44: 1416e24.](#)

[32] [Zhang Q, Ma H, Mai K, Zhang W, Liufu Z, Xu W. Interaction of dietary *Bacillus subtilis* and fructooligosaccharide on the growth performance, non-specific immunity of sea cucumber, *Apostichopus japonicus*. Fish Shellfish Immunol 2010;29\(2\):204e11.](#)

[33] [Sun Y, Wen Z, Li X, Meng N, Mi R, Li Y, et al. Dietary supplement of fructooligosaccharides and *Bacillus subtilis* enhances the growth rate and](#)

[disease resistance of the sea cucumber *Apostichopus janonicus* \(Selenka\). *Aquacult Res* 2012;43:1328e34.](#)

[34] [Sohn KS, Kim MK, Kim JD, Han IK. The role of immunostimulants in mono gastric animal and fish-review. *Asian Austral J Anim* 2000;13\(8\):1178e87.](#)

[35] [Linehan SA, Martínez-Pomares L, Gordon S. Macrophage lectins in host defence. *Microbes Infect* 2000;2\(3\):279e88.](#)

[36] [Benites V, Gilharry R, Gernat AG, Murillo JG. Effect of dietary mannan oligo saccharide from Bio-Mos or SAF-Mannan on live performance of broiler chickens. *J Appl Poult Res* 2008;17\(4\):471e5.](#)

[37] [Klebaniuk R, Matras J, Patkowski K, Picta M. Effectiveness of Bio-MOS in sheep nutrition. *Annu Anim Sci* 2008;8:369e80.](#)

[38] [Yang Y, Iji PA, Choct M. Dietary modulation of gut microflora in broiler chickens: a review of the role of six kinds of alternatives to in-feed antibiotics. *World's Poult Sci J* 2009;65:97e114.](#)

[39] [Torrecillas S, Montero D, Izquierdo M. Improved health and growth of fish fed mannan oligosaccharides: potential mode of action. *Fish Shellfish Immunol* 2014;36:525e44.](#)

[40] [Staykov Y, Spring P, Denev S, Sweetman J. Effect of a mannan oligosaccharide on the growth performance and immune status of rainbow trout \(*Oncorhynchus mykiss*\). *Aquacult Int* 2007;15\(2\):153e61.](#)

[41] [Rodriguez-Estrada U, Satoh S, Haga Y, Fushimi H, Sweetman J. Effects of single and combined supplementation of *Enterococcus faecalis*, mannan oligosaccharide and polyhydrobutyric acid on growth performance and immune response of rainbow trout *Oncorhynchus mykiss*. *Aquacult Sci* 2009;54\(4\): 609e17.](#)

[42] [Rodriguez-Estrada U, Satoh S, Haga Y, Fushimi H, Sweetman J. Effects of inactivated *Enterococcus faecalis* and mannan oligosaccharide and their combination on growth, immunity, and disease protection in rainbow trout. *N Am J Aquacult* 2013;75:416e28.](#)

[43] [Samrongpan C, Areechon N, Yoonpundh R, Sirsapoome P. Effect of mannan oligosaccharides on growth, survival and disease resistance of Nile tilapia \(*Oreochromis niloticus* Linnaeus\) fry. In: 8th international symposium on tilapia in aquaculture; October 12e14, 2008. pp. 345e53. Cairo, Egypt.](#)

[44] [Torrecillas S, Makol A, Benítez-Santana T, Caballero MJ, Montero D, Sweetman J, et al. Reduced gut bacterial translocation in European sea bass \(*Dicentrarchus labrax*\) fed mannan oligosaccharides \(MOS\). *Fish Shellfish Immunol* 2011;30\(2\):674e81.](#)

[45] [Groff J, LaPatra S. Infectious diseases impacting the commercial culture of salmonids. *J Appl Aquac* 2000;10:17e90.](#)

[46] [Birkbeck TH, Ringø E. Pathogenesis and the gastrointestinal tract of growing fish. In: Holzapfel W, Naughton P, editors. *Microbial ecology in growing animals*. Edinburgh, UK: Elsevier; 2005. pp. 208e34.](#)

[47] [Ringø E, Myklebust R, Mayhew TM, Olsen RE. Bacterial translocation and pathogenesis in the digestive tract of larvae and fry. *Aquaculture* 2007;268: 251e64.](#)

[48] [Ringø E, Løvmo L, Kristiansen M, Salinas I, Myklebust R, Olsen RE, et al. Lactic acid bacteria vs. pathogens in the gastrointestinal tract of fish: a review. *Aquacult Res* 2010;41:451e67.](#)

- [49] [Sang HM, Ky LT, Fotedar R. Dietary supplementation of mannan oligosaccharide improves the immune responses and survival of marron, *Cherax tenuimanus* \(Smith, 1912\) when challenged with different stressors. *Fish Shellfish Immunol* 2009;27\(2\):341e8.](#)
- [50] [Sang HM, Fotedar R, Filer K. Effects of dietary mannan oligosaccharide on survival, growth, physiological condition, and immunological responses of marron. *Cherax tenuimanus* \(Smith 1912\). *J World Aquacult Soc* 2011;42:230e41.](#)
- [51] [Sang HM, Fotedakar R, Filer K. Effects of dietary mannan oligosaccharide on the survival, growth, immunity and digestive enzyme activity of freshwater crayfish, *Cherax destructor* Clark \(1936\). *Aquacult Nutr* 2011;17:e629e35.](#)
- [52] [Sang HM, Fotedar R. Effects of mannan oligosaccharide dietary supplementation on performances of the tropical spiny lobsters juvenile \(*Panulirus ornatus*, Fabricius 1798\). *Fish Shellfish Immunol* 2010;28\(3\):483e9.](#)
- [53] [Gu M, Ma H, Mai K, Zhang W, Bai N, Wang X. Effects of dietary \$\beta\$ -glucan, mannan oligosaccharide and their combinations on growth performance, immunity and resistance against *Vibrio splendidus* of sea cucumber, *Apostichopus japonicus*. *Fish Shellfish Immunol* 2011;31:303e9.](#)
- [54] [Bai N, Zhang W, mai K, Gu M, Xu W. Effects of continuous and alternative administration of \$\alpha\$ -glucan and mannan-oligosaccharide on the growth, immunity and resistance against *Vibrio splendidus* of sea cucumber *Apostichopus japonicus* \(Selenka\). *Aquacult Res* 2013;44:1613e24.](#)
- [55] [Ebrahimi Gh, Ouraji H, Khalesi MK, Sudagar M, Barari A, Zarei Dangesaraki M, et al. Effects of a prebiotic, Immunogen[®], on feed utilization, body composition, immunity and resistance to *Aeromonas hydrophila* infection in the common carp *Cyprinus carpio* \(Linnaeus\) fingerlings. *J Anim Physiol Anim Nutr* 2012;96:591e9.](#)
- [56] [Yang ST, Silva EM. Novel products and new technologies for use of a familiar carbohydrate, milk lactose. *J Dairy Sci* 1995;78\(11\):2541e62.](#)
- [57] [Sako T, Matsumoto K, Tanaka R. Recent progress on research and applications of non-digestible galacto-oligosaccharides. *Int Dairy J* 1999;9\(1\):69e80.](#)
- [58] [Vos AP, M'rabet L, Stahl B, Boehm G, Garssen J. Immune-modulatory effects and potential working mechanisms of orally applied nondigestible carbohydrates. *Crit Rev Immunol* 2007;27\(2\):97e140.](#)
- [59] [Torres DPM, Goncalves M do Pilar F, Teixeira JA, Rodrigues LR. Galacto-oligosaccharides: production, properties, applications, and significance as prebiotics. *Compr Rev Food Sci F* 2010;9:438e54.](#)
- [60] [Zhou QC, Buentello JA, Gatlin III DM. Effects of dietary prebiotics on growth performance, immune response and intestinal morphology of red drum \(*Sciaenops ocellatus*\). *Aquaculture* 2010;309:253e7.](#)
- [61] [Grootaert C, Delcour JA, Courtin CM, Broekaert WF, Verstraete W, Van de Wiele T. Microbial metabolism and prebiotic potency of arabinoxylan oligo-saccharides in the human intestine. *Trends Food Sci Tech* 2007;18:64e71.](#)

- [62] [Swennen MK, Courtin CM, Lindemans GCJE, Delcour JA. Large-scale production and characterization of wheat bran arabinoxyloligosaccharides. J Sci Food Agric 2006;86:1722e31.](#)
- [63] [Izydorczyk MS, Biliaderis Costas. Cereal arabinoxylans: advances in structure and physicochemical properties. Carbohydr Polym 1995;28:33e48.](#)
- [64] [Geralylou Z, Souffreau C, Rurangwa E, D'Hondt S, Callewaert L, Courtin CM, et al. Effects of arabionxylan-oligosaccharides \(AXOS\) on juvenile Siberian sturgeon \(*Acipenser baerii*\) performance, immune responses and gastrointestinal microbial community. Fish Shellfish Immunol 2012;33:718e24.](#)
- [65] [Geralylou Z, Souffreau C, Rurangwa E, De Meester L, Courtin CM, Delcour JA, et al. Effects of dietary arabionxylan-oligosaccharides \(AXOS\) and endogenous probiotics on the growth performance, non-specific immunity and gut microbiota of juvenile Siberian sturgeon \(*Acipenser baerii*\). Fish Shellfish Immunol 2013;35:766e75.](#)
- [66] [Ringø E, Zhou Z, Olsen RE, Song SK. Use of chitin and krill in aquaculture e the effect on gut microbiota and the immune system: a review. Aquacult Nutr 2012;18:117e31.](#)
- [67] [Lam KL, Cheung PCK. Non-digestible long chain beta-glucans as novel pre biotics. Bioact Carbohydr Diet Fibre 2013;2:45e64.](#)
- [68] [Chiu ST, Tsai RT, Hsu JP, Liu CH, Cheng W. Dietary sodium alginate administration to enhance the non-specific immune responses, and disease resistance of the juvenile grouper *Epinephelus fuscoguttatus*. Aquaculture 2008;277: 66e72.](#)
- [69] [Geng X, Dong XH, Tan BP, Yang QH, Chi SY, Liu HY, et al. Effects of dietary chitosan and *Bacillus subtilis* on the growth performance, non-specific immunity and disease resistance of cobia, *Rachycentron canadum*. Fish Shellfish Immunol 2011;31:400e6.](#)
- [70] [Harikrishnan R, Kim MC, Kim JS, Han YJ, Jang IS, Balasundaram C, et al. Immunomodulatory effect of sodium alginate enriched diet in kelp grouper *Epinephelus brneus* against *Streptococcus iniae*. Fish Shellfish Immunol 2011;30:543e9.](#)
- [71] [Harikrishnan R, Kim JS, Balasundaram C, Heo MS. Immunomodulatory effects of chitin and chitosan enriched diets in *Epinephelus bruneus* against *Vibrio alginolyticus* infection. Aquaculture 2012;326e329:46e52.](#)
- [72] [Ibrahem MD, Fathi M, Meslhy S, El-Aty AMA. Effect of dietary supplementation of inulin and vitamin C on the growth hematology, innate immunity and resistance of Nile tilapia \(*Oreochromis niloticus*\). Fish Shellfish Immunol 2010;29:241e6.](#)
- [73] [Ahmdifar E, Akrami R, Ghelichi A, Zarejabad AM. Effects of different dietary prebiotic inulin levels on blood serum enzymes, hematologic, and biochemical parameters of great sturgeon \(*Huso huso*\) juveniles. Comp Clin Path 2011;20: 447e51.](#)
- [74] [Cerezuela R, Cuesta A, Mesequer J, Esteban MA. Effects of inulin on gilthead seabream \(*Sparus aurat L.*\) innate immune parameters. Fish Shellfish Immunol 2008;24:663e8.](#)
- [75] [Cerezuela R, Guardiola FA, Mesequer J, Esteban MA. Increases in immune parameters by inulin and *Bacilius subtilis* dietary administration to gilthead seabream \(*Sparus aurata L.*\) did not correlate with disease](#)

[resistance to Photobacterium damsela. Fish Shellfish Immunol 2012;32\(6\):1032e40.](#)

[76] [Cerezuela R, Cuesta A, Meseguer J, Esteban MA. Effects of inulin and heat-activated Bacillus subtilis on gilthead seabream \(Sparus aurata L.\) innate immune parameters. Benef Microbes 2012;3:77e81.](#)

[77] [Mourino JLP, Do Nascimento Vieira F, Jatobá AB, Da Silva BC, Jesus GFA, Seiffert WQ, et al. Effect of dietary supplementation of inulin and W. cibaria on haemato-immunological parameters of hybrid surubim \(Pseudoplatystoma sp\). Aquacult Nutr 2012;18:73e80.](#)

[78] [Brown GD, Gordon S. Immune recognition: a new receptor for beta glucans. Nature 2001;413:36e7.](#)

[79] [Kudrenko B, Snape N, Barnes AC. Linear and branched b\(1e3\) D-glucans activate but do not prime teleost macrophages in vitro and are inactivated by dilute acid: implications for dietary immunostimulation. Fish Shellfish Immunol 2009;26:443e50.](#)

[80] [Ringø E, Olsen RE, Vecino JLG, Wadsworth S, Song SK. Use of immunostimulants and nucleotides in aquaculture: a review. J Mar Sci Res Dev 2012;1:104.](#)

[81] [Dhanasiri AKS, Brunvold L, Brinchmann MF, Korenes K, Bergh Ø, Kiron V. Changes in the intestinal microbiota of wild Atlantic cod Gadus morhua L. upon captive rearing. Microb Ecol 2011;61:20e30.](#)

[82] [Galli C, Calder PC. Effects of fat and fatty acid intake on inflammatory and immune responses. A critical review. Annu Nutr Metab 2009;55:123e39.](#)

[83] [Cerezuela R, Meseguer J, Esteban MA. current knowledge in synbiotic use for fish aquaculture. A review. J Aquacult Res Dev 2011;\(S1\):008.](#)

[84] [Ganguly S, Paul I, Mukhopadhyay SK. Application and effectiveness of immunostimulants, probiotics, and prebiotics in aquaculture: a review. Israeli J Aquacult e Bamidgah 2010;62:130e8.](#)

[85] [Shoelson SE, Herrero L, Naaz A. Obesity, inflammation, and insulin resistance. Gastroenterology 2007;132\(6\):2169e80.](#)

[86] [de Heredia FP, Gomez-Martínez S, Macrós A. Obesity, inflammation and the immune system. Proc Nutr Soc 2012;71\(2\):332e8.](#)

[87] [Gomez GD, Balcazar JL. A review on the interactions between gut microbiota and innate immunity of fish. FEMS Immunol Med Microbiol 2008;52:145e54.](#)

[88] [Perez T, Balcazar JL, Ruiz-Zarzuola I, Halalhel N, Vendrell D, de Blas I, et al. Host-microbiota interactions within the fish intestinal ecosystem. Mucosal Immunol 2010;3:355e60.](#)